

Instruction for use

EIA CMV IgA

REF CMA096



Kit for professional use

IVD **CE**₁₀₂₃

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Enzyme immunoassay for the detection of IgA antibodies to Cytomegalovirus in human serum, plasma or cerebrospinal fluid

1 Introduction

Human cytomegalovirus (CMV, Human Herpesvirus 5, HHV 5) is a member of the *Herpetoviridae* family.

Primary infection of CMV occurs mainly in childhood or adolescence. The infection can be transmitted in different ways (e.g. respiratory apparatus, urogenital tract). Clinically, the disease process is usually asymptomatic or mild (fever, fatigue, mononucleosis symptoms). Dormant infections where the virus survives can be reactivated – generally by changes in host-virus relations (pregnancy, serious disease, stress or immunosuppressive treatment). Reinfection is possible by a different strain of CMV. CMV infection during pregnancy causes developmental defects (virus is easily transmitted through the placenta and affects the foetus). During primary infection mother-to-infant transmission of the virus (i.e. via placenta) occurs in 1/3 to 1/2 of cases; however, during the reactivation stage, transplacental transmission occurs in only 1% of cases.

Diagnosis of the disease is based on a clinical picture and laboratory tests. The most widespread serological method used for the detection of specific IgA, IgM and IgG (avidity) antibodies to CMV in laboratory diagnosis of the infection is ELISA.

IgA

Antibodies of IgA class are a sign of an active infection – primary infection as well as reactivation. The IgA antibodies are produced during reactivation. Their presence may be, but often is not, accompanied by presence of specific IgM antibodies. Specific IgA antibodies are very important for confirmation of CMV infection reactivation when they are present together with IgG antibodies.

IgM

Production of IgM antibodies usually increases a few weeks after infection and then (during 4–6 months) decreases slowly. In immunosuppressed patients, the IgM antibodies can be present at low levels even two years after infection. As IgM antibodies can also be produced during reactivation, IgM determination alone cannot discriminate primary infection from reactivation.

IgG

Specific IgG antibodies can be detected approx. 1 week after the increase of IgM and IgA antibodies. Their seroconversion (increase of titre) indicates primary infection. The method of IgG avidity detection is used for discrimination between primary infection and reactivation. It is important for the risk assessment of congenital transmission.

Detection of IgG antibodies to CMV is also used as a standard method for screening at blood donors.

2 Test Principle

The kit is intended for detection of specific IgA antibodies in a sample by means of a sandwich type of the EIA method (i.e. a solid phase coated with specific antigen – antibody from the analysed sample – labelled antibody). The labelled antibody (conjugate) is an animal immunoglobulin fraction to human IgA conjugated with horseradish peroxidase. Peroxidase activity is determined in the test by a substrate containing TMB. Positivity is indicated when blue colour appears; after stopping solution has been added, blue changes to yellow. The yellow colour intensity is measured by a photometer at 450 nm, and it is proportional to the concentration of specific IgA antibodies in the sample.

Antigen Used

Purified and inactivated antigen isolated from CMV AD 169 strain with a high content of specific immunodominant epitopes

3 Materials Provided

MICROPLATE	Microtitre Plate coated with antigen, 12 x 8 wells in bag with desiccant	1 pc
CONTROL -	Negative Control Solution containing no specific human antibodies, ready to use	1 × 2 ml
CUTOFF	CUT-OFF Solution containing specific human antibodies in cut-off concentration, ready to use	1 × 3 ml
CONTROL +	Positive Control Solution containing specific human antibodies, ready to use	1 × 2 ml
CONJUGATE	Conjugate Solution containing peroxidase labelled animal immunoglobulin to human IgA, ready to use	1 × 15 ml
DILUENT 2	Sample Diluent 2 Buffer with protein stabilisers, ready to use	1 × 105 ml
SUBSTRATE 2	TMB-Complete 2 Chromogenic substrate solution containing TMB/H ₂ O ₂ , ready to use	1 × 15 ml
WASH 20x	Wash Solution 20× concentrated buffer	1 × 75 ml
STOP	Stop Solution Acid solution, ready to use	1 × 15 ml
	Instructions for use	1 pc

4 Other Material Required for Manual Test Performance

Single and multichannel pipettes

Disposable tips

Microplate washer

Timer

Incubator (37°C)

Microplate reader

5 Storage and Stability

Store the kit at +2°C to +8°C. Do not freeze. If the kit is stored as described, the labelled expiration date is valid. The expiration date is indicated on the package. The opened kit should be used within three months.

Samples Preparation and Storage

The following human body liquids can be used for testing: serum, citrate plasma and cerebrospinal fluid. Anticoagulants in the plasma (except for citrate) as well as bacterially contaminated, haemolytic or chylous samples can affect the test results.

Samples can be stored at +2°C to +8°C for one week. For a longer period, store samples at -20°C. Diluted samples should be used as soon as possible.

6 Preparation of Reagents

Dilute the Wash Solution 1:20 (1 part of solution and 19 parts of distilled water); e.g. 75 ml of the concentrated Wash Solution + 1425 ml of distilled water.

Salt crystals might develop in the bottle with the concentrated Wash Solution. Prior to use, it is necessary to dissolve the crystals by warming the bottle in a water bath. The diluted Wash Solution is stable at +2°C to +8°C for one week.

The Controls (positive, negative and CUT-OFF) are ready to use, do not dilute further!

The Conjugate is ready to use, do not dilute further!

TMB-Complete is a one-component chromogenic substrate solution ready to use, do not dilute further!

Interchangeability of reagents

The Sample Diluent, TMB-Complete and the Avidity Solution are interchangeable in EIA kits of TestLine Clinical Diagnostics s.r.o., provided they have the identical numeric marking (e.g. Sample Diluent 2, Sample Diluent 3, etc.). The Stop Solution and the Wash Solution are universal in all kits.

7 Preparation of Samples

Mix gently the Sample Diluent prior to use.

Dilution of sera and plasma samples

Dilute well mixed samples 1:101 with the Sample Diluent:

E.g.: 10 µl of sample + 1 ml of the Sample Diluent

Mix well.

Dilution of cerebrospinal fluid samples (CSF)

Dilute well mixed CSF 1:3 with the Sample Diluent:

E.g.: 50 µl of CSF + 100 µl of the Sample Diluent

Mix well.

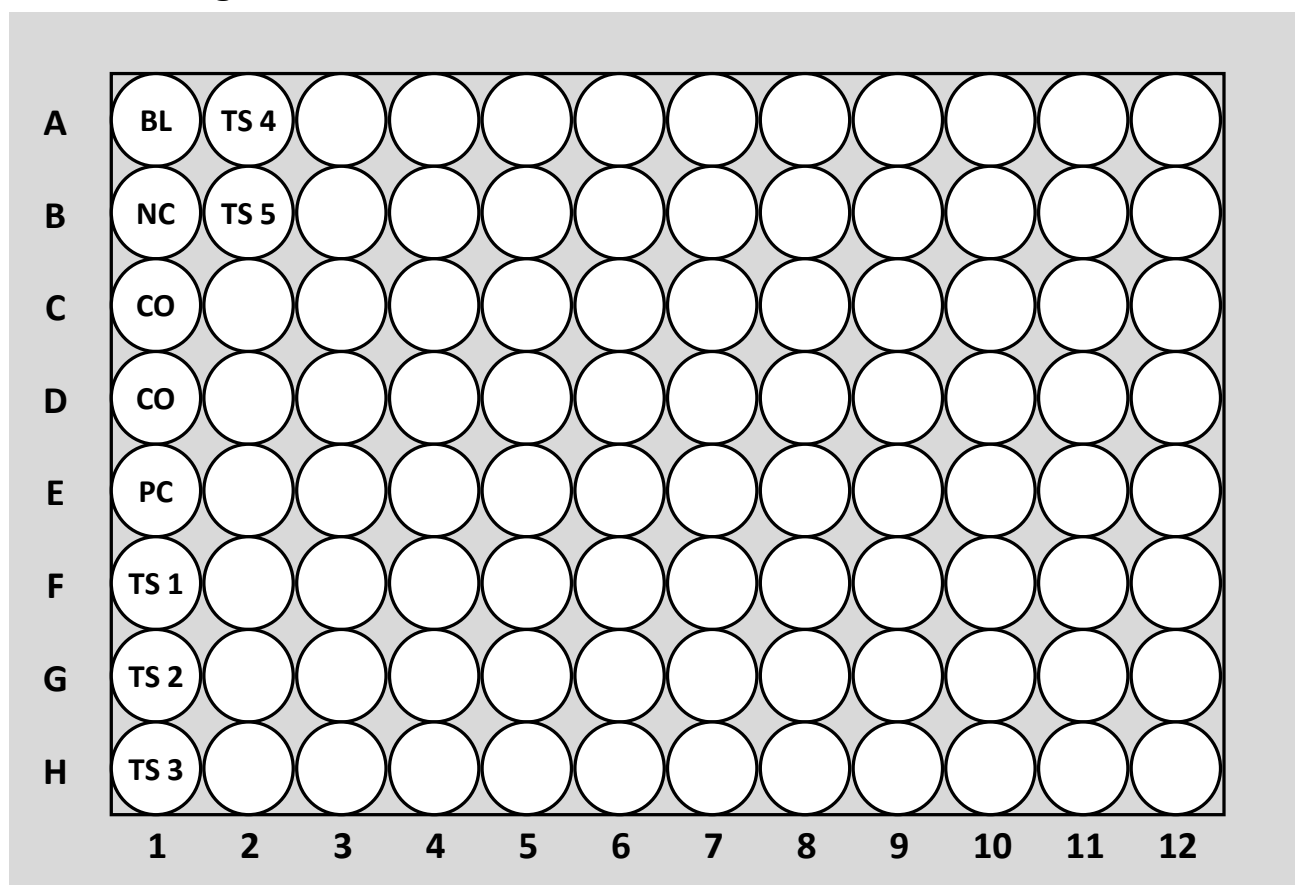
8 Assay Procedure

Allow all reagents to come to room temperature and mix well. If you do not use a whole microplate, return unnecessary strips into the bag with desiccant. Seal the bag tightly and store at +2°C to +8°C. Keep dry!

1. Dispense the controls and the diluted samples according to the working schedule.
 - Leave A1 well empty (blank).
 - Pipette 100 µl of the Negative Control into 1 well.
 - Pipette 100 µl of CUT-OFF into 2 wells.
 - Pipette 100 µl of the Positive Control into 1 well.
 - Pipette 100 µl of the diluted samples (see Chapter Preparation of Samples) into the other wells.
2. Cover the microplate with the lid and incubate at 37°C for 30 minutes.
3. Aspirate the content of the wells and wash 5× with the working strength Wash Solution. Fill the wells up to the edge. Finally, tap the inverted microplate thoroughly on an absorbent paper to remove solution remnants.
4. Pipette 100 µl of the Conjugate into all wells except A1 well.
5. Cover the microplate with the lid and incubate at 37°C for 30 minutes.
6. Aspirate the content of the wells and wash 5× with the working strength Wash Solution. Fill the wells up to the edge. Finally, tap the inverted microplate thoroughly on an absorbent paper to remove solution remnants.
7. Pipette 100 µl of TMB-Complete into all wells. Avoid contamination – see Chapter Procedural Notes.
8. Cover the microplate with the lid and incubate at 37°C for 30 minutes. Keep out of light.

9. Stop the reaction by adding 100 μ l of the Stop Solution in the same order and intervals as the substrate was added.
10. Read the colour intensity in wells against blank (A1 well) using photometer set to 450 nm. The absorbance should be read within 30 minutes after stopping the reaction.

9 Working Schedule



BL	Blank (empty well)
NC	100 μ l <input type="text" value="CONTROL -"/>
CO	100 μ l <input type="text" value="CUTOFF"/>
PC	100 μ l <input type="text" value="CONTROL +"/>
TS 1-x	100 μ l diluted tested sample

10 Quality Control

The test is valid if:

The absorbance of blank is lower than 0.150.

$$\text{BLANK} < 0.150$$

The absorbance of the Negative Control is lower than half of the mean absorbance of CUT-OFF.

$$\text{CONTROL}^- < 0.5 \times \text{CUTOFF}$$

The mean absorbance of CUT-OFF is within a range of 0.150 – 0.900.

$$0.150 < \text{CUTOFF} < 0.900$$

The absorbance of the Positive Control is higher than the mean absorbance of CUT-OFF.

$$\text{CONTROL}^+ > \text{CUTOFF}$$

11 Results Interpretation

Calculation of Index of Positivity (IP)

Divide the absorbance of a tested sample by the mean absorbance of CUT-OFF measured in the same test run:

$$\text{IP} = \frac{\text{Absorbance of sample}}{\text{Mean absorbance of CUT-OFF}}$$

Interpretation of the test results is described in Table 1.

Table 1 Interpretation of results

Index of Positivity (IP)	Evaluation
lower than 0.9	negative
0.9 to 1.1	borderline
higher than 1.1	positive

Examination of borderline samples, i.e. samples with Index of Positivity from 0.9 to 1.1, should be repeated from a new sample collected after 2 to 6 weeks regarding to the disease specificities.

Serological finding can be interpreted only in the context of results of other laboratory tests and patient clinical picture.

12 Safety Precautions

The kit is intended for in vitro diagnostic use only.

The sera used for controls were tested and found to be negative for HIV 1 and HIV 2, HBsAg, HCV, TPHA. In spite of this fact, they still need to be handled as potentially infectious materials.

Some reagents contain sodium azide, which is a toxic compound. Avoid contact with skin.

The Stop Solution contains diluted acid solution. Avoid contact with eyes and skin.

It is necessary to observe the local safety rules and regulations.

First aid

In case of contact with eyes, flush with copious amount of water and seek medical assistance. In case of contact with skin and clothing, remove all the contaminated clothes. Wash the skin with soap and plenty of running water. In case of contact with solutions containing plasma or clinical samples, disinfect the skin. In case of accidental ingestion, flush the mouth with drinking water and seek medical assistance.

Remnants disposal

All the materials used for performing the test must be treated as potentially infectious due to the contact with biological materials. Therefore they need to be disposed together with biological waste.

Expired kit disposal

Disassemble the kit and dispose the components as biological material. Discard the packaging material as required by local regulations.

13 Procedural Notes

In order to obtain reliable results, it is necessary to **strictly follow the Instructions for Use**. Always use clean preferably disposable tips and glassware.

Microtitre Plate – in order to prevent water condensation on the surface of the microplate, always allow the bag with the microplate to warm up to room temperature before opening.

Wash Solution – use high quality distilled water for preparing the working strength Wash Solution.

Washing procedure – keep to the prescribed number of wash cycles and fill the wells to the upper edge. The soak time (i.e. interval between two different wash cycles during which the wells stay filled up with the Wash Solution) should be approx. 30-60 seconds.

TMB-Complete – the vessel used for multichannel pipetting should not be used for other reagents. Do not return the surplus TMB-Complete from the pipetting vessel into the vial.

Non-reproducible results might be caused by improper methodology as following:

- insufficient mixing of reagents and samples before use
- improper replacement of vial caps
- using the same tip for pipetting different reagents
- reagent exposure to excessive temperature; bacterial or chemical contamination
- insufficient washing or filling of the wells (the wells should be filled to the upper edge), improper aspiration of Wash Solution remnants
- contamination of the well edges with Conjugate or samples
- using reagents from different kit lots
- contact of reagents with oxidants, heavy metals and their salts

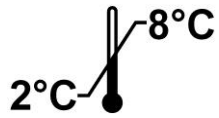
The kit might be used for sequential examinations. When preparing working strength solutions, use only the amount of reagents needed for the analysis.

The kit might be used in all types of automatic EIA analysers.

If necessary, TestLine Clinical Diagnostics s.r.o. can offer a certified modification of the Instructions for Use for the specific type of analyser.

The producer cannot guarantee that the kit will function properly if the assay procedure instructions are not strictly adhered to.

14 IFU Symbols



Temperature limitation



Keep dry



Expiry date



Lot number



Manufactured by



Consult instructions



Catalogue number



Number of tests














In vitro diagnostic medical device

Notes

Notes

Notes

Summary of EIA CMV IgA Protocol

Step No.	Symbol	Test steps
1		Dilute samples serum/plasma 1:101 (10 µl + 1 ml) cerebrospinal fluids 1:3 (50 µl + 100 µl)
2		Pipette Controls and diluted samples – 100 µl Blank = empty well
3		Incubate at 37°C for 30 mins
4		Aspirate and wash the wells 5×
5		Pipette Conjugate – 100 µl Blank = empty well
6		Incubate at 37°C for 30 min
7		Aspirate and wash the wells 5×
8		Pipette Substrate (TMB-Complete) – 100 µl Including blank
9		Incubate at 37°C for 30 min
10		Pipette Stop Solution – 100 µl Including blank
11		Read colour intensity at 450 nm